

Effect of C2/C3-*endo* Unsaturation on the Cytotoxicity and DNA-Binding Reactivity of Pyrrolo[2,1-*c*][1,4]benzodiazepines

Stephen J. Gregson,^{a,†} Philip W. Howard,^{a,†} Simona Barcella,^a Anthonia Nakamya,^a
Terence C. Jenkins,^b Lloyd R. Kelland^c and David E. Thurston^{a,*,†}

^aCRC Gene Targeted Drug Design Research Group, School of Pharmacy and Biomedical Science, University of Portsmouth,
St Michael's Building, White Swan Road, Portsmouth, Hants PO1 2DT, UK

^bYorkshire Cancer Research Laboratory of Drug Design, Cancer Research Group, University of Bradford, Bradford,
West Yorkshire BD7 1DP, UK

^cCRC Centre for Cancer Therapeutics, Institute for Cancer Research, Clifton Avenue, Sutton, Surrey SM2 5PX, UK

Received 4 May 2000; revised 19 June 2000; accepted 20 June 2000

Abstract—A series of novel C2/C3-*endo* unsaturated pyrrolo[2,1-*c*][1,4]benzodiazepines (PBDs) has been synthesised via cleavage of the N10-Alloc protecting group from appropriate precursors. Biophysical and biological evaluations show that the presence of C2/C3-*endo* unsaturation in the PBD C-ring enhances both DNA-binding reactivity and in vitro cytotoxic potency. © 2000 Elsevier Science Ltd. All rights reserved.

The pyrrolo[2,1-*c*][1,4]benzodiazepines (PBDs) are a class of antitumour antibiotics known to interact covalently in the minor groove of DNA in a sequence-selective manner.¹ Covalent bonding occurs between the C11 position of the PBD and the exocyclic N2 group of guanine, giving rise to a preference for purine–guanine–purine sequences.¹ Of the two best known natural products, anthramycin (**1**) and tomaymycin (**2**) (Fig. 1), the former binds to DNA more efficiently², a characteristic thought to be associated with the C2/C3-*endo* unsaturation of anthramycin.

Although structure–activity relationship (SAR) information regarding both A-ring substitution³ and C-ring C2-*exo* unsaturation^{2,4} is available, little is known about the effect of varying C2-substituents in C-ring C2/C3-*endo* unsaturated PBDs. Here we report the synthesis and biophysical and biological evaluation of five novel C2/C3-*endo* unsaturated PBDs with unique C2-substituents not found in PBD natural products isolated to date.

The C2/C3 unsaturation under investigation was introduced by Horner–Emmons olefination of ketones **4a,b** (Scheme 1).⁴ In the presence of excess ylid (2.2 equiv),

spontaneous migration of the initially formed *exo* double bond into the ring occurred to give the C2/C3-*endo* unsaturated compounds **5a–c** in high yields. A similar bond migration process in related molecules was observed by both Kaneko and Leimgruber.^{5,6} TBAF was found to be incompatible with amides **5a–c**, so the TBDMS protecting groups were instead removed by treatment with AcOH/THF/H₂O to provide alcohols **6a–c** in high yields.⁷ B-ring closure was accomplished by oxidation under Swern conditions⁸ to afford **7a–c** in good yield. Further elaboration of the C-ring involved LiBH₄ reduction of ester **5b** to provide the homoallylic alcohol **8** (55% yield), followed by acetylation under standard conditions to give the acetate **9** in 58% yield (Scheme 2).

Selective cleavage of the TBDMS ether proceeded under mild conditions to furnish the primary alcohol **10** in 81% yield. Spontaneous oxidation/cyclisation under Swern conditions gave the Alloc-protected carbinolamine **11** (82%) which was hydrolysed in aqueous K₂CO₃ to afford the PBD precursor **12** in 78% yield. The *N*-protected carbinolamines (**7a–c**, **11**, **12**) were converted to the novel PBD derivatives **13–17**^{9,10} in good yields using the method of Deziel¹¹ (Scheme 3).

The data presented in Table 1 show that the five C2/C3-*endo* unsaturated PBDs **13–17** exhibit sub-micromolar IC₅₀ values in a panel of human ovarian tumour cell

*Corresponding author. Fax: +44-115-951-3114; e-mail: david.thurston@nottingham.ac.uk

[†]Present address: Cancer Research Laboratories, School of Pharmaceutical Sciences, University of Nottingham, Nottingham, NG7 2RD, UK.

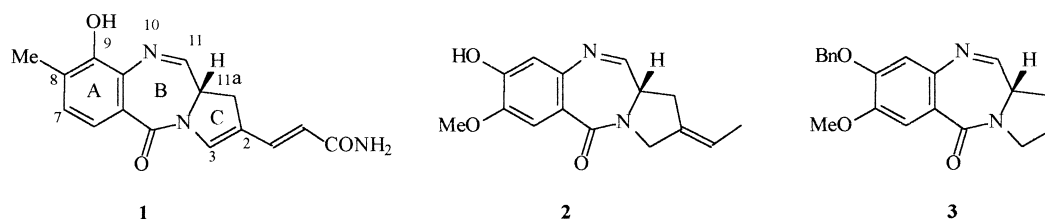
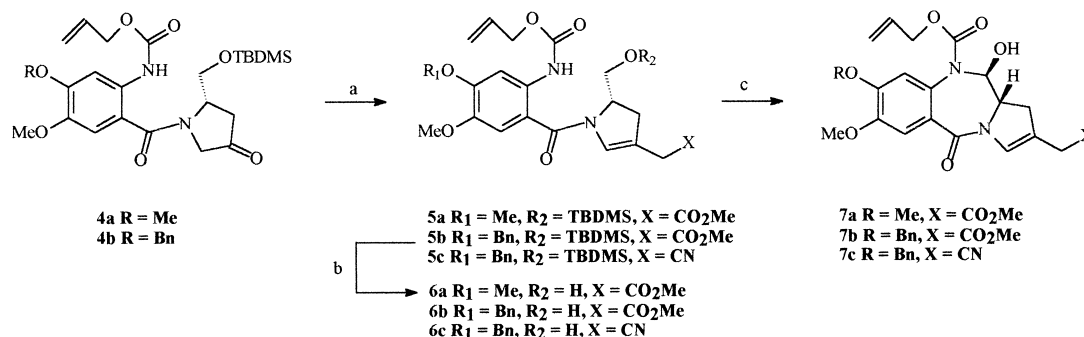


Figure 1.

Table 1. In vitro cytotoxicity (human ovarian) and thermal denaturation data (calf thymus DNA) for the novel C2-*endo* unsaturated PBDs **13–17** and parent compounds **1** and **3**

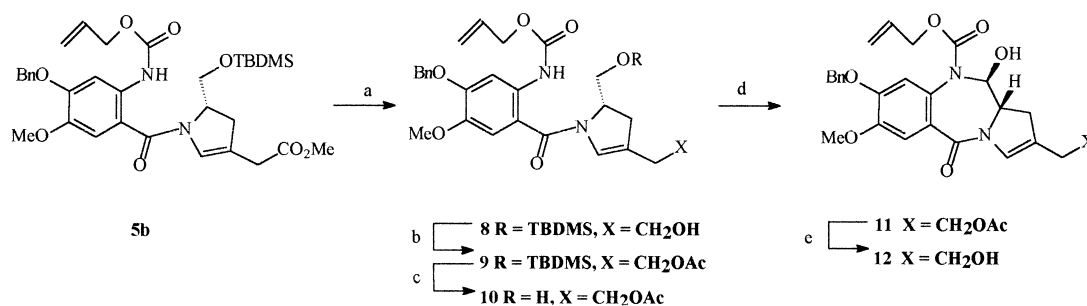
Compound no.	Cytotoxicity (μM) ^a							Induced ΔT_m ($^{\circ}\text{C}$) ^c after incubation at 37°C for:		
	SKOV-3	A2780	A2780 ^{cisR}	RF ^b	CH1	CH1 ^{cisR}	RF ^b	0 h	4 h	18 h
13	0.45	0.36	0.46	1.3	0.115	0.15	1.3	3.41	4.63	5.94 ^d
14	0.12	0.0145	0.12	8.3	0.016	0.04	2.5	3.08	3.96	4.70 ^d
15	0.52	0.155	0.43	2.8	0.105	0.27	2.6	1.84	2.31	3.48 ^d
16	0.46	0.10	0.27	2.7	0.105	0.16	1.5	1.57	2.11	2.53
17	0.105	0.07	0.105	1.5	0.09	0.037	0.4	3.98	4.61	5.60
1	0.16	0.155	0.160	1.03	0.062	0.05	0.81	9.40	11.20	13.0
3	0.46	0.17	0.48	2.8	0.145	0.145	1.0	0.20	0.40	0.40

^aDose of PBD required to inhibit cell growth by 50% compared with PBD-free controls after incubation for 96 h at 37°C .^bRF = resistance factor (IC_{50} cisplatin-resistant/parent).^cFor a 5:1 molar ratio of duplex CT-DNA ($100\mu\text{M}$) and ligand ($20\mu\text{M}$) in aqueous buffer (10mM $\text{NaH}_2\text{PO}_4/\text{Na}_2\text{HPO}_4 + 1\text{mM}$ Na_2EDTA , pH 7.00 ± 0.01). All values are $\pm < 0.09$ – 0.13°C .^d ΔT_m values of 6.65, 4.97 and 4.31°C (all $\pm < 0.19^{\circ}\text{C}$) were determined for PBDs **13**, **14** and **15**, respectively, after incubation for 36 h.**Scheme 1.** (a) $(\text{C}_2\text{H}_5\text{O})_2\text{P}(\text{O})\text{CH}_2\text{CO}_2\text{CH}_3$, NaH, THF, 0°C , 73% for **5a**, 80% for **5b**; $(\text{C}_2\text{H}_5\text{O})_2\text{P}(\text{O})\text{CH}_2\text{CN}$, NaH, THF, 0°C , 63% for **5c**; (b) AcOH:THF:H₂O (3:1:1), 98% for **6a**, 90% for **6b**, quant for **6c**; (c) $(\text{COCl})_2$, DMSO, TEA, CH_2Cl_2 , -45°C , 54% for **7a**, 66% for **7b**, 68% for **7c**.

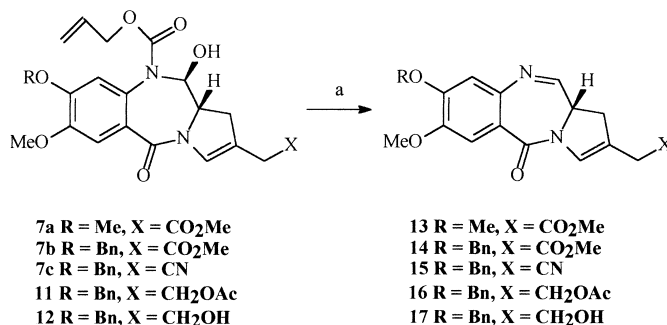
lines.¹² In this particular assay, the homoallylic alcohol PBD **17** and the methyl ester **14** are the most potent compounds. Unusually, **17** had a resistance factor of 0.4 in the CH1/CH1^{cisR} pair, indicating that this compound has more activity in the CH1^{cisR} cell line than the parent line suggesting that it may have potential in the treatment of cisplatin-resistant disease. The compounds were also evaluated for their ability to raise the melting temperature (T_m) of calf thymus (CT) DNA.² The data illustrate that the C2/C3-*endo* unsaturated PBDs have a greater ability to stabilise CT-DNA than compounds with fully saturated C-rings or C2-*exo*-unsaturation.^{3,4} There is a broad correlation between DNA stabilisation and cytotoxicity, with **14** and **17** having two of the highest ΔT_m values. However, **13** gave the highest ΔT_m of all the synthetic compounds which may reflect the

presence of the smaller C8-methoxy substituent. Interestingly, the novel PBDs are of similar cytotoxic potency to the natural product anthramycin (**1**), although the latter elevates the DNA melting temperature by a far greater extent. These results suggest that the relationship between T_m and cytotoxicity may not be as straightforward as previously thought.^{1–3}

In summary, the ketone intermediates **4a,b** are ideal substrates for the introduction of C2/C3-*endo* unsaturation into the C-ring of PBDs. Compared to C8-*O*-benzyl DC-81 which has a saturated and unsubstituted C-ring (**3**), the enhanced DNA binding affinity and sub-micromolar cytotoxicity observed for **13–17** illustrates the significant effect that C-ring unsaturation has on both biological activity and biophysical properties.



Scheme 2. (a) LiBH₄, THF, 0°C, 55%; (b) (Ac)₂O, pyridine, 25°C, 58%; (c) AcOH:THF:H₂O (3:1:1), 81%; (d) (COCl)₂, DMSO, TEA, CH₂Cl₂, -45°C, 82%; (e) K₂CO₃, MeOH, CH₂Cl₂, 78%.



Scheme 3. (a) Pd(PPh₃)₄, PPh₃, pyrrolidine, CH₂Cl₂, quant (13–17).

Acknowledgements

The Cancer Research Campaign (UK) is thanked for providing financial support of this work (SP1938/0301 to D.E.T./T.C.J., and SP1938/0401 to DET). Additional support was provided by Yorkshire Cancer Research (to T.C.J.).

References and Notes

- Thurston, D. E. In *Molecular Aspects of Anticancer Drug–DNA Interactions*; Neidle, S.; Waring, M. J., Eds.; Macmillan Press: UK, 1993; Vol. 1, p 54.
- Puvvada, M. S.; Hartley, J. A.; Jenkins, T. C.; Thurston, D. E. *Nucleic Acids Res.* **1993**, *21*, 3671.
- Thurston, D. E.; Bose, D. S.; Howard, P. W.; Jenkins, T. C.; Leoni, A.; Baraldi, P. G.; Guiotto, A.; Cacciari, B.; Kelland, L. R.; Foloppe, M.-P.; Rault, S. *J. Med. Chem.* **1999**, *42*, 1951.
- Gregson, S. J.; Howard, P. W.; Corcoran, K. E.; Barcella, S.; Yasin, M. M.; Hurst, A. A.; Jenkins, T. C.; Kelland, L. R.; Thurston, D. E. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 1845.
- Kaneko, T.; Wong, H.; Doyle, T. W.; Rose, W. C.; Bradner, W. T. *J. Med. Chem.* **1985**, *28*, 388.
- Leimgruber, W.; Batcho, A. D.; Czajkowski, R. C. *J. Am. Chem. Soc.* **1968**, *90*, 5641.
- Marshall, J. A.; Sedrani, R. *J. Org. Chem.* **1991**, *56*, 5496.
- Fukuyama, T.; Liu, G.; Linton, S. D.; Lin, S.-C.; Nishino, H. *Tetrahedron Lett.* **1993**, *34*, 2577.
- The C8-OH form of nitrile **15** has been reported previously by Tozuka and co-workers: Tozuka, Z.; Yazawa, H.; Murata, M.; Takaya, T. *J. Antibiot.* **1983**, *36*, 1699.
- Data for **16**: [α]_D²¹ = +741.67° (c=0.66, CHCl₃); ¹H NMR (270 MHz, CDCl₃) δ 7.78 (d, 1H, J=4.03 Hz), 7.52 (s, 1H), 7.46–7.28 (m, 5H), 6.83 (s, 1H), 6.82 (s, 1H), 5.21–5.20 (m, 2H), 4.29–4.16 (m, 2H), 3.96 (s, 3H), 3.88–3.87 (m, 1H), 3.33–3.16 (m, 1H), 3.05–2.98 (m, 1H), 2.53–2.48 (m, 2H), 2.07 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃) (rotamers) δ 170.9, 162.6, 161.1, 150.9, 148.2, 140.1, 136.1, 132.1, 132.0, 128.7, 128.6, 128.1, 127.3, 124.7, 121.4, 111.9, 111.6, 70.8, 61.9, 56.2, 53.6, 37.4, 27.9, 21.0; MS (EI), m/z (relative intensity) 420 ([M]⁺, 14), 419 (12), 418 (36), 360 (20), 328 (3), 313 (8), 267 (22), 256 (4), 129 (3), 105 (3), 91 (100), 65 (5); IR (CHCl₃) 3313 (br), 2957, 2934, 1736, 1598, 1509, 1455, 1437, 1384, 1243, 1179, 1120, 1096, 1037, 697, 542 cm⁻¹; exact mass calcd for C₂₄H₂₄N₂O₅ m/z 420.1685, obsd m/z 420.1750.
- Deziel, R. *Tetrahedron Lett.* **1987**, *28*, 4371.
- Kelland, L. R.; Abel, G.; McKeage, M. J.; Jones, M.; Goddard, P. M.; Vallenti, M.; Murrer, B.; Harrap, K. R. *Cancer Res.* **1993**, *53*, 2581.